

# Observations on Southern White Rhinoceros *Ceratotherium simum simum* Translocated to Uganda

Tropical Conservation Science  
Volume 11: 1–7  
© The Author(s) 2018  
Article reuse guidelines:  
sagepub.com/journals-permissions  
DOI: 10.1177/1940082918806805  
journals.sagepub.com/home/trc



Douglas Sheil<sup>1</sup>  and Andrew E. Kirkby<sup>2</sup>

## Abstract

Translocation is increasingly common in conservation but remains controversial due, in part, to limited information. Here, in a brief study, we consider the translocation of the southern white rhinoceros *Ceratotherium simum simum* to a fenced 64-km<sup>2</sup> reserve in Uganda (a replacement for the locally extinct native northern subspecies, *Ceratotherium simum cottoni*). We examined the condition, behavior, and habitat use of two female-calf groups. Rhinos were least active during the early afternoon. Both groups spent more time feeding in *Microchloa kunthii* grasslands than expected given the small extent of such areas but spent most time in the more abundant woodlands and *Hyparrhenia* grasslands. The small ranges of the two groups, 11.8 and 9.0 km<sup>2</sup> (100 h of observations over a 1-month period in 2011), and their apparent good health suggest satisfactory feeding conditions. Reassuringly, we identify no concerns but also note that our judgments are constrained by limited information and guidance. There is scope for novel observations, for example, our observations suggest that mothers lead calves more frequently in habitats with reduced visibility versus more open habitats. Every translocation is an experiment, but lessons will be lost without greater emphasis on transparency and assessments.

## Keywords

evidence-based conservation, ethology, translocation, reintroduction, habitat use

## Introduction

Translocations for conservation involve the transfer of living organisms from one area to another (see, e.g., International Union for Conservation of Nature [IUCN], 2013). The intention is to augment, establish, or reestablish a population (Schwartz & Martin, 2013; Seddon, Griffiths, Soorae, & Armstrong, 2014). Although such translocations are increasingly common their costs, risks and uncertainties make them controversial (Minteer & Collins, 2010; Müller & Eriksson, 2013; Ricciardi & Simberloff, 2009; Schwartz & Martin, 2013). These controversies result, in part, from limited information and transparency. Good information is often unavailable. Observations on survival and reproduction appear insufficient if the strange and neurotic behaviors that are sometimes seen in captive animals might occur in translocated animals (Mason & Latham, 2004; Rose, Nash, & Riley, 2017; Wolfensohn et al., 2018). Studies are an essential element to evaluate efforts and improve evidence and available information (Rout, Hauser, & Possingham, 2009; Sutherland et al., 2013).

Translocations are a key strategy for the conservation of rhinoceros species (Emslie, Amin, & Kock, 2009). The IUCN guidelines for such interventions note that many past rhino translocations have lacked monitoring and that “as a result nothing or little was learned to help guide and improve future translocations and the translocations have often ended in failure” (Emslie et al., 2009). Here, we examine white rhinos (*Ceratotherium simum* Burchell, 1817) translocated to a protected ranch in Uganda.

<sup>1</sup>Faculty of Environmental Sciences and Natural Resource Management, Norwegian University of Life Sciences, Ås, Norway

<sup>2</sup>Wildlife Conservation Society, Villa Ifanomezantsoa, Face II A 78 D Soavimbahoaka, Antananarivo, Madagascar

Received 31 July 2018; Revised 15 September 2018; Accepted 23 September 2018

### Corresponding Author:

Douglas Sheil, Faculty of Environmental Sciences and Natural Resource Management, Norwegian University of Life Sciences, 1430 Ås, Norway.  
Email: douglas.sheil@nmbu.no



Uganda lost its last white rhinoceros to poaching in 1982 (Edroma, 1982). In 1997, the nongovernmental organization “Rhino Fund Uganda” was founded with the aim of establishing white rhinos in Uganda (Brett, 2002). The northern subspecies previously occurred in Uganda, but with these animals unavailable, the southern subspecies was judged a suitable replacement (Brett, 2002). The translocation aims to bolster tourism and spread the risk to this threatened species. Furthermore, rhinos can maintain habitats, including wallows and short grasslands, that benefit other species (see, e.g., Everatt, Andresen, Ripple, & Kerley, 2016): For example, wildebeest and impala have been observed to leave areas once rhinos were absent (Waldram, Bond, & Stock, 2008).

IUCN lists the white rhinoceros as “Near Threatened” but notes “the continued and increased poaching threat” (<http://www.iucnredlist.org>; for a more extensive review of the threats, see Taylor et al., 2017). The northern subspecies, *C. simum* ssp. *cottoni*, is “Critically Endangered (Possibly Extinct in the Wild)” (<http://www.iucnredlist.org>). The southern subspecies, *C. simum* ssp. *simum*, has fared better. Despite the fact that in 1962 there was only a single wild population which comprised fewer than 50 animals, by 2010 captive breeding and translocation had resulted in over 20,000 animals (Emslie et al., 2009; Emslie, Milliken, & Talukdar, 2012). Some consider the northern and southern white rhinoceros as distinct species (*C. cottoni* and *C. simum*, respectively; Groves, Fernando, & Robovský, 2010; but see also Harley, de Waal, Murray, & O’Ryan, 2016). If distinct, the establishment of the southern form in Uganda is not a “reintroduction,” as it was not previously present. Furthermore, the two rhinos live in different parts of Africa, and it has been speculated that they may have different habitat preferences (Groves et al., 2010). For example, Southern White Rhinos are known to feed on short, nutritious grasses, whereas Northern White Rhinos typically lived in wetter habitats and may have eaten more dicotyledons and tall grasses (Forbes-Watson, 1967).

In 2002, an area in central Uganda was selected as suitable for white rhinos due to its “short grassland in open and wooded areas” and a predicted ability to support at least 20 animals (see Brett, 2002). A 64.2-km<sup>2</sup> fenced ranch was established in 2004 and is now called “Ziwa Rhino Sanctuary” (hereafter “Ziwa,” Figure 1, inset). Six rhinos were introduced: four from the Solio Ranch in Kenya and two from the Disney Animal Kingdom, USA (Patton, 2009). Since 2009, each adult female has calved regularly resulting in 18 offspring (7 females and 11 males) by June 2018. Details are available in studbooks, reports, and websites (see <http://rhinoresourcecenter.com> and <http://www.rhinofund.org>). Aspects of social organization of the animals at

Ziwa are described in Patton, Campbell, and Genade (2018); in brief, adult males tend to be solitary while adult females associate with their youngest offspring and sometimes with adolescent animals that may be unrelated.

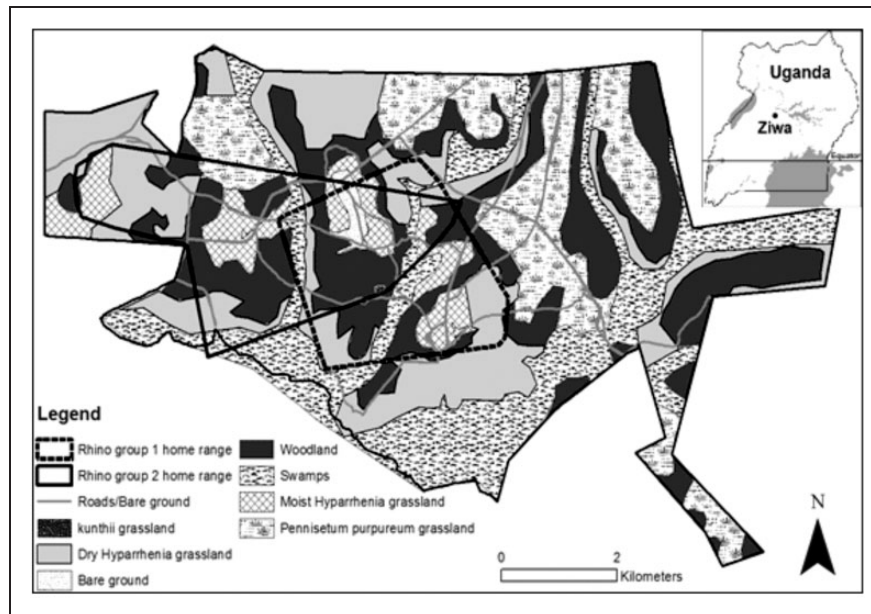
Translocations of white rhinos have not always had successful outcomes (McKeown, 2016; Player, 1967). For example, 21 of 59 animals translocated from Natal, South Africa, to Maputo, Mozambique, between 1967 and 1970 were dead by 1973. Five died from the translocation itself (anesthesia and stress), one was killed by another rhino prior to release, and two were found dead soon after. Seven more died after becoming trapped in swamps, three more drowned, and neighboring communities killed another three. Of the 38 remaining, many had left the reserve and some were believed to have returned to Natal (Tello, 1973 cited in McKeown, 2016). Others have noted behavioral problems in translocated animals, for example, only grazing at night (Player, 1967). It is because of such experiences that the IUCN guidelines on rhino translocation state that “Animals should be extensively monitored in the immediate post-release settling in period to determine the extent of their movements/dispersal and use of habitat (water points, feeding areas, and habitat types) at different times since release and in the different seasons. This information and maps of kernel-estimated home ranges can help refine and improve best-practice release recommendations” (Emslie et al., 2009, p. 99).

Here, we examine the state and behavior of two adult female-led groups 6 years after their introduction at Ziwa. Our goal was to contribute toward achieving these IUCN guidelines and help evaluate how the animals had adapted to local conditions.

## Methods

### Study Site

Ziwa is situated in Nakasongola, in central Uganda at 1°26′53.50″N and 32° 4′40.10″E (Figure 1, inset top right). The ranch is relatively flat averaging 1,045 m a.s.l., but two steeper hills rise to 1,082 and 1,080 m a.s.l. in the eastern corners. Annual rainfall averages about 1,000 mm (Brett, 2002; Patton, Campbell, Genade, Ayiko, & Lutalo, 2011). Regional vegetation includes woodlands, grasslands, and wetlands. Although tsetse (*Glossina* spp.) occur, trypanosome infections are not considered a threat. Large carnivores are largely absent though there is believed to be one leopard (*Panthera pardus pardus*) that visits the ranch where small prey are relatively common (e.g., Reedbuck *Redunca redunca*, Bushbuck *Tragelaphus scriptus*, Ugandan Kob *Kobus kob thomasi*, Common duiker *Sylvicapra grimmia*, and Vervet monkeys *Chlorocebus pygerythrus*).



**Figure 1.** Ziwa, location, vegetation, and range of the white rhino groups. The ranch is situated in Nakasongola, in central Uganda at  $1^{\circ}26'53.50''N$  and  $32^{\circ}4'40.10''E$  (inset top right). The map was produced through a habitat survey (15 m by 15 m plots,  $n = 421$ ), conducted over the reserve combined with visual discrimination using recent Google Earth satellite raster imagery from a viewing altitude of 3.6 km. The boundary on the south western side is a permanent wetland swamp with no fence, causing variations in the calculations of the total area of the reserve. Seven habitat types were identified, including five Food and Agriculture Organization grassland types (Mwebaze, 1999) here by label (area and percent of total) description: Woodland (16.6 km<sup>2</sup>, 27.6%)—trees over 3 m. Common species include *Combretum molle*, *Acacia siberiana*, *Bauhinia thonningii*, and shrubby *Scutia myrtina*, and *Rhus* spp.; Swamp (13.9 km<sup>2</sup>, 23.1%)—permanently and temporarily flooded areas over 30 cm deep. Plants include rushes, sedges, and *Hyparrhenia* spp. grasses. Trees include Palms; Moist *Hyparrhenia* grassland (4.0 km<sup>2</sup>, 6.6%)—dominated by *Brachiaria* spp and *Cynodon nlemfuensis* and typically including: *Hyparrhenia rufa*, *Hyparrhenia variabilis*, *Chloris gayana*, *Panicum maximum*, and *Imperata cylindrica*; Dry *Hyparrhenia* grassland (14.3 km<sup>2</sup>, 23.8%)—dominated by *Hyparrhenia dissoluta* and including *Setaria sphacelata*, *Themeda triandra*, *Cenchrus ciliaris*, *Cynodon nlemfuensis*, *Andropogon gayanus*, *Andropogon schriensis*, and *Heteropogon contortus*; *Microchloa kunthii* grassland (0.04 km<sup>2</sup>, 0.07%)—rocky with shallow soils, dominated by *M. kunthii*; *Pennisetum purpureum* grassland (10.1 km<sup>2</sup>, 16.7%)—coarse, tall grassland dominated by *P. purpureum*; often > 2 m high. Occurs in wetter fertile areas; Bare ground/roads (1.3 km<sup>2</sup>, 2.1%)—no vegetation over 5 m<sup>2</sup> or more.

### Field Methods

Two groups (“Group 1” with two animals and “Group 2” with three animals) were monitored on alternating days, for 50 h each during the months of July and August 2011. Both groups included a 10-year-old adult female (“Bella” and “Kori” both born in 2000 in Solio Ranch, Kenya, and arrived at Ziwa together in July 2005, n.b. neither mother was pregnant at the time of our observations) with their first calves both 2-year-old males (Bella’s calf was born on June 24, 2009, and Kori’s on January 2, 2010). At the time of the study, Group 2 also included an unrelated 2-year-old male.

Rhinos were individually identified using ear notches and horn shapes (Støen, Pitlagano, & Moe, 2009). The animals are guarded and are thus habituated; nonetheless, monitoring at night was judged impractical, as the animals sometimes react aggressively to torchlight (rangers note that the animals tend to linger in specific locations at night where they generally rest and feed, see

also Patton et al., 2011). At the start of the study, the body condition of the two adult females was scored (following Adcock & Emslie, 2003).

We recorded the behavior using a continuous sampling approach. Each morning, groups were located and followed for approximately 10 h from a distance of around 20 m (further if animals appeared agitated). The animals were followed from behind or to the side but never from in front. In the case of a charge, protocol was to rapidly find a safe location (this never occurred during the study but had been reported previously). We noted time, location, habitat, and temperature when behaviors changed (using a handheld GPS and a “Weathereye WEA25” located at 1.3 m). We established 397 15-m × 15-m vegetation plots while monitoring and added 24 randomly located habitat survey points across the rest of the reserve. In each plot, we measured grass height, identified the dominant species, and applied standard Food and Agriculture Organization classifications (Mwebaze, 1999).

Habitat preferences were analyzed using compositional analysis where relative habitat use (time) and availability (area) are ranked against each other (Aebischer, Marcstrom, Kenward, & Karlbom, 1993; Aitchison, 1986). The total area used by the animals was estimated from the Minimum Convex Polygon from our GPS points ( $n=395$ ) using Hawth's tools in ArcGIS 9.3. Statistical tests were performed in SPSS 19 (<https://www.ibm.com/products/spss-statistics>).

## Results

Both adult females scored 4 out of 5 on the Adcock and Emslie (2003)'s condition scores—indicating above average health for females with calves.

We distinguished and mapped seven habitat types and found that three—Woodland, Dry *Hyparrhenia* grassland, and Swamp—comprised well over half of the total cover (with 27.6, 23.8, and 23.1%, respectively, see Figure 1).

The areas used by Groups 1 and 2 were 9 km<sup>2</sup> and 12 km<sup>2</sup>, respectively (Figure 1). The predominant activities were grazing, resting, and moving (45%, 39%, and 10% of observation time, respectively). Episodes of resting averaged 30 min, while grazing and moving averaged 12 and 8 min each. Other behaviors comprised less than 6% of the total time (Table 1). Both groups occasionally wallowed after drinking in hot conditions (average temperature 32°C).

Play was mainly play-fighting between juveniles or a juvenile and visiting male. The mother would watch any play-fighting and quickly intervened if the juvenile became upset—roaring and even charging to drive the other animal away (six episodes).

Most periods of movement were short (average 8 min), as animals shifted grazing locations. The proportion of time spent moving for both groups was relatively constant throughout the day (standard deviations for the percentage of observations were 12.8 and 8.9 for Groups 1 and 2, respectively; by comparison, standard deviations for resting were 32.7 and 28.7). Both groups were most active between 07:30 a.m. and 09:30 a.m., and between 3:00 p.m. and 5:30 p.m. Resting was favored from 10:00 a.m. until 02:30 p.m. (Figure 2(a)). On a typical day, temperatures rose until 01:00 p.m. (reaching about 30°C) and declined after around 04:30 p.m. Hence, rhinos generally rested during the warmest hours but resumed activity before afternoon temperatures dropped (Figure 2(a)). Group 2 was more active than Group 1, spending 35.3% versus 44.5% of total observations at rest. Temperature was positively correlated with time spent grazing and resting and negatively correlated with moving ( $p = .014, .008, <.001$ , respectively,  $n = 16$ , Kendall's Tau-b). We also found that animals spent less time moving in areas with shorter versus longer grasses ( $p < .001$ ).

Juveniles sometimes lead activities. For example, they would often whimper at some point in the mornings, prompting the mother to stop grazing and lead the way to a resting place. The juvenile would sometimes end the resting period too by getting up, grazing, and vocalizing, until its mother also got up. Moving groups too were sometimes lead by juveniles, but we noted that adults lead more frequently in denser versus more open vegetation (Mann-Whitney  $U = 89$ ,  $p = .042$ ,  $N = 42$ , 32 with mother leading and 10 juvenile leading, with a mean local tree density of 12.9 and 6.4 stems per 15-m  $\times$  15-m plot, respectively).

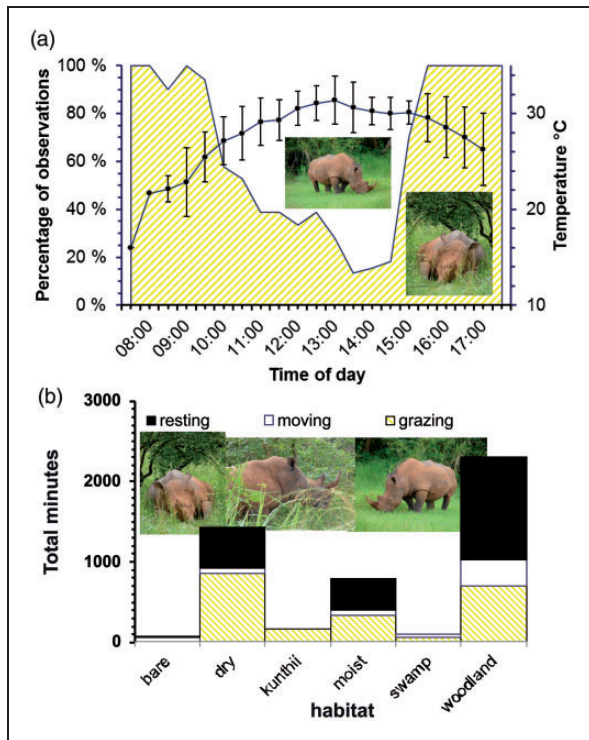
**Table 1.** Summary of Observed Activities by Time and Episodes.

Activities	Total minutes	Total episodes	Average episode (min)	Longest episode (min)	Shortest episode (min) <sup>a</sup>
Grazing	2,501	203	12.3	70	1
Resting	2,169	72	30.1	90	2
Moving	571	71	8.0	45	1
Playing	90	11	8.0	20	1
Drinking	72	8	9.0	20	3
Clay licking <sup>b</sup>	55	3	18.3	30	5
Wallowing	39	9	4.3	22	1
Fighting	24	6	6.5	13	1
Total	5,553	382	14.5	90	1

*Grazing* is recorded when animals are actively feeding on vegetation; *Resting* when the head and body are on the ground; *Moving* involves walking head up facing forward (as movement and grazing were often combined, this is only recorded when animals stopped feeding for over half a minute); *Playing* involved the young animals interacting with others; *Drinking* involves animals having their head down at water; *Clay licking* is self-explanatory; *Fighting* is recorded when the adult female actively attempts to drive off another animal (play fighting is included under playing); and *Wallowing* involves scratching at the ground in the water hole or lying down and rolling.

<sup>a</sup>The minimum period recorded is 1 min.

<sup>b</sup>Clay licking was only observed for one group at one location on one day.



**Figure 2.** Activity for both rhino groups combined (a) by time of day for all observations (shaded area,  $n = 212$  observations for Rhino Group 1,  $n = 189$  for Rhino Group 2) along with average temperatures (with standard deviation  $n = 401$ ) and (b) minutes of principle activity by habitat type (Rhino Group 1,  $n = 197$ , intervals of observation where behavior was constant, Group 2,  $n = 164$ ). All photographs by A. Kirkby.

Both rhino groups spent most time in Woodland, followed by Dry, then Moist *Hyparrhenia* grassland (Figure 2(b)). Both groups had similar preferences among the seven habitat categories (Kendall's Tau- $b = 0.683$ ,  $p = .048$ ): favoring *M. kunthii* grassland and woodlands and avoiding *P. purpureum* grassland (bare ground being a discrepancy ranked third for Group 1 and avoided entirely by Group 2).

## Discussion

Our goal was to assess the degree to which southern white rhinos were adapted to conditions at Ziwa. Despite its limitations, our study complements and corroborates studies based on ranger records (Patton, 2009; Patton et al., 2018). The animals are healthy with no obvious indications of unusual or pathological behaviors—though our ability to make such assessments is constrained by available information.

At Ziwa, the animals spent most time feeding in the relatively extensive woodland and *Hyparrhenia* grasslands. Similar behaviors have been seen in South Africa, where woodlands fulfilled most requirements

for shade and palatable grasses (Pienaar, 1994a). The Ziwa animals avoided steep slopes and the tall *P. purpureum* grasslands (we know that captive animals are sometimes fed on this grass; Droma, 2012). That both groups of animals fed in the small areas of *M. kunthii* more frequently than expected by chance suggests that this is a preferred habitat. Animals also appear to linger where the grass is shorter. The Ziwa animals are known to aggregate on young grass after burning (Patton et al., 2018). Reports from southern Africa confirm that shorter greener grasses tend to be favored by grazing rhinos (Adcock & Emslie, 2003; Jordaan, 2010; Pienaar, 1994a; Shrader, Owen-Smith, & Ogutu, 2006; Støen et al., 2009).

Overall, though comparative information is limited, the animals' behaviors did not appear out of the ordinary for the species (see also Patton et al., 2011). Overlapping ranges are normal (e.g., Rachlow, Kie, & Berger, 1999), and the area used by each group lies within the annual ranges seen in southern Africa though much larger areas are unexceptional (Pedersen, 2009; Pienaar, 1994b; Rachlow et al., 1999). Some variation likely reflects different methods, but the small area implies adequate food availability too. A short study like this cannot provide a complete picture: likely, the area used would increase over a full year of observations, as animals are reported to favor higher ground after good rains (Patton et al., 2011). There again, even a year of data cannot account for intra-annual variation or requirements following droughts or other rare events—so all such data need to be seen as provisional. Activity patterns were similar to those in southern Africa (Jordaan, 2010; Shrader et al., 2006; Støen et al., 2009). We saw no signs that the climate caused problems though the animals often rested within woodland shade through the hottest periods each day and appeared to drink and to wallow more frequently in warmer conditions.

One lesson is how our ability to judge normal behaviors is limited by suitable reference information. Despite valuable compilations on aspects of rhino biology (e.g., Ververs et al., 2017), there is no simple means to compare and judge the behavior of these animals.

## Implications for Conservation

Conservation has not always learned as much from its failures as it might have, and transparency and information sharing are not always prioritized (for a more detailed discussion, see, e.g., Bower et al., 2018; Meek et al., 2015; Sutherland et al., 2013). In the case of Ziwa's rhinos, our ability to judge "normal" behaviors was constrained by available information. These limitations also allowed us to identify apparently undocumented behaviors: while we know mother rhinos are protective (Hutchins & Kreger, 2006), we found no previous mention that they permit

juveniles to lead only when visibility is good. We need more information on all aspects of behavior.

Our observations raise no concerns about the planned Ziwa translocation, though more study is clearly required (Emslie et al., 2009). Wherever the animals are ultimately released must provide water, shade, and short grasses. Monitoring will remain crucial. The threat of poaching means that the habituated animals will require guarding. In this context, any monitoring by the guards will be valuable. We encourage studies that complement these approaches. We need to know more about translocations and their outcomes.

### Acknowledgments

We are grateful to Rhino Fund Uganda and Ziwa Ranch, specifically Angie Genade for supporting A. E. K. during his MSc research. A. E. K. thanks the rangers of Ziwa for their kindness and help and Dr. David Harper for his guidance.

### Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

### Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

### ORCID iD

Douglas Sheil  <http://orcid.org/0000-0002-1166-6591>

### References

- Adcock, K., & Emslie, R. (2003). *Monitoring African Rhino—'Sandwiches' training course for field rangers* (5th ed., pp. 1–116). Harare, Zimbabwe: SADC Regional Programme for Rhino Conservation.
- Aebischer, N. J., Marcstrom, V., Kenward, R. E., & Karlbom, M. (1993). *Survival and habitat utilisation: A case for compositional analysis*. Basel, Switzerland: Birkhaeuser Verlag.
- Aitchison, J. (1986). *The statistical analysis of compositional data*. New York, NY: Chapman and Hall.
- Bower, S. D., Brownscombe, J. W., Birnie, -Gauvin, K., Ford, M. I., Moraga, A. D., Bennett, J. R. (2018). Making tough choices: Picking the appropriate conservation decision-making tool. *Conservation Letters*, 11(2), e12418.
- Brett, R. (2002). *Feasibility study for the establishment of a rhino sanctuary for black and white rhinos at Ziwa Ranch, Buruli, Uganda*. Retrieved from Harare, SADC Regional Programme for Rhino Conservation: <http://www.rhinoresourcecenter.com>
- Droma, A. (2012). The monitoring of Rhino Health Through Bleeding at the Uganda Wildlife Education Centre In: *Proceedings of the International Conference of Zookeepers (presented at the 4th International congress on Zoo Keeping in Singapore. 9–13 September, 2012*. Retrieved from [http://www.rhinoresourcecenter.com/pdf\\_files/134/1347678879.pdf](http://www.rhinoresourcecenter.com/pdf_files/134/1347678879.pdf).
- Edroma, E. L. (1982). White rhino extinct in Uganda. *Oryx*, 16(04), 352–357.
- Emslie, R., Amin, R., & Kock, R. (Eds.). (2009). *Guidelines for the in situ reintroduction and translocation of African and Asian rhinoceros*. Gland, Switzerland: International Union for Conservation of Nature.
- Emslie, R., Milliken, T., & Talukdar, B. (2012). *African and Asian rhinoceroses—Status, conservation and trade*. Gland, Switzerland: International Union for Conservation of Nature.
- Everatt, K. T., Andresen, L., Ripple, W. J., & Kerley, G. I. (2016). Rhino poaching may cause atypical trophic cascades. *Frontiers in Ecology and the Environment*, 14(2), 65–67.
- Forbes-Watson, A. D. (1967). The square-lipped rhino (*Ceratotherium simum cottoni* (Lydekker)) in Uganda. *East African Wildlife*, 5, 167–171.
- Groves, C. P., Fernando, P., & Robovský, J. (2010). The sixth rhino: A taxonomic re-assessment of the critically endangered northern white rhinoceros. *PLOS One*, 5(4), e9703.
- Harley, E. H., de Waal, M., Murray, S., & O'Ryan, C. (2016). Comparison of whole mitochondrial genome sequences of northern and southern white rhinoceroses (*Ceratotherium simum*): The conservation consequences of species definitions. *Conservation Genetics*, 17(6), 1285–1291. doi:10.1007/s10592-016-0861-2
- Hutchins, M., & Kreger, M. (2006). Rhinoceros behaviour: Implications for captive management and conservation. *International Zoo Yearbook*, 40(1), 150–173.
- International Union for Conservation of Nature. (2013). *Guidelines for reintroductions and other conservation translocations*. Gland, Switzerland: Author.
- Jordaan, H. (2010). *Behavioural-ecology of the white rhinoceros (Ceratotherium simum) in the Willem Pretorius Game Reserve* (Masters of Science, University of South Africa, Pretoria). Retrieved from <http://hdl.handle.net/10500/3593>
- Mason, G. J., & Latham, N. (2004). Can't stop, won't stop: Is stereotypy a reliable animal welfare indicator? *Animal Welfare*, 13, S57–S69.
- McKeown, K. (2016). Rehoming rhinos in southern Africa: Animal indigeneity and wildlife translocations in the 1960s and 1970s. *Critical African Studies*, 1–20.
- Meek, M. H., Wells, C., Tomalty, K. M., Ashander, J., Cole, E. M., Gille, D. A., & Yamane, L. (2015). Fear of failure in conservation: The problem and potential solutions to aid conservation of extremely small populations. *Biological Conservation*, 184, 209–217.
- Minteer, B. A., & Collins, J. P. (2010). Move it or lose it? The ecological ethics of relocating species under climate change. *Ecological Applications*, 20(7), 1801–1804.
- Müller, H., & Eriksson, O. (2013). A pragmatic and utilitarian view of species translocation as a tool in conservation biology. *Biodiversity Conservation*, 22, 1837–1841.
- Mwebaze, S. M. N. (Ed.). (1999). *Country pasture/forage resource profiles: Uganda*. Rome, Italy: Food and Agriculture Organization of the United Nations.
- Patton, F. J. (2009). Bringing rhinos back to Uganda. *Swara*, 4, 46–49.

- Patton, F. J., Campbell, P. E., & Genade, A. (2018). The development of White Rhino social organisation at Ziwa Rhino Sanctuary. *Pachyderm*, *57*, 112–113.
- Patton, F. J., Campbell, P. E., Genade, A., Ayiko, R., & Lutalo, G. (2011). The behaviour of white rhinos at Ziwa Rhino Sanctuary, Uganda with particular reference to night-time activity. *Pachyderm*, *50*, 77–83.
- Pedersen, G. (2009). *Habitat use and diet selection of reintroduced white rhinoceros (Ceratotherium simum) in Pafuri, Kruger National Park* (MSc Thesis, Stellenbosch University, Stellenbosch, South Africa). Retrieved from <http://scholar.sun.ac.za/handle/10019.1/4037>
- Pienaar, D. (1994a). *Habitat preferences of the white rhino in the Kruger National Park*. Paper presented at the Proceedings of Symposium on “Rhinos as Game Ranch Animals”. Onderstepoort, Republic of South Africa, 9–10 September 1994.
- Pienaar, D. (1994b). *Social organization and behaviour of the white rhinoceros*. Paper presented at the Proceedings of Symposium on “Rhinos as Game Ranch Animals”. Onderstepoort, Republic of South Africa, 9–10 September 1994.
- Player, I. (1967). *The translocation of the white rhinoceros a success in wild life conservation in Southern Africa*. Paper presented at the The Role of Zoos in International Conservation, San Diego, USA.
- Rachlow, J. L., Kie, J. G., & Berger, J. (1999). Territoriality and spatial patterns of white rhinoceros in Matobo National Park, Zimbabwe. *African Journal of Ecology*, *37*(3), 295–304.
- Ricciardi, A., & Simberloff, D. (2009). Assisted colonization is not a viable conservation strategy. *Trends in Ecology and Evolution*, *24*(5), 248–253.
- Rose, P. E., Nash, S. M., & Riley, L. M. (2017). To pace or not to pace? A review of what abnormal repetitive behavior tells us about zoo animal management. *Journal of Veterinary Behavior: Clinical Applications and Research*, *20*, 11–21. doi:10.1016/j.jveb.2017.02.007
- Rout, T. M., Hauser, C. E., & Possingham, H. P. (2009). Optimal adaptive management for the translocation of a threatened species. *Ecological Applications*, *19*(2), 515–526.
- Schwartz, M. W., & Martin, T. G. (2013). Translocation of imperiled species under changing climates. *Annals of the New York Academy of Sciences*, *1286*(1), 15–28.
- Seddon, P. J., Griffiths, C. J., Soorae, P. S., & Armstrong, D. P. (2014). Reversing defaunation: Restoring species in a changing world. *Science*, *345*(6195), 406–412.
- Shrader, A. M., Owen-Smith, N., & Ogutu, J. O. (2006). How a mega-grazer copes with the dry season: Food and nutrient intake rates by white rhinoceros in the wild. *Functional Ecology*, *20*(2), 376–384. doi:10.1111/j.1365-2435.2006.01107.x
- Støen, O. G., Pitlagano, M. L., & Moe, S. R. (2009). Same-site multiple releases of translocated white rhinoceroses *Ceratotherium simum* may increase the risk of unwanted dispersal. *Oryx*, *43*(4), 580–585. doi:10.1017/s0030605309990202
- Sutherland, W. J., Mitchell, R., Walsh, J., Amano, A., Ausden, M., Beebee, T. J. C., Dicks, L. V. (2013). Conservation practice could benefit from routine testing and publication of management outcomes. *Conservation Evidence*, *10*, 1–3.
- Taylor, A., Balfour, D., Brebner, D. K., Coetzee, R., Davies-Mostert, H., Lindsey, P. A., t Sas-Rolfes, M. (2017). Sustainable rhino horn production at the pointy end of the rhino horn trade debate. *Biological Conservation*, *216*, 60–68. doi:10.1016/j.biocon.2017.10.004
- Ververs, C., van Zijll Langhout, M., Hostens, M., Otto, M., Govaere, J., Durrant, B., & Van Soom, A. (2017). Reproductive performance parameters in a large population of game-ranched white rhinoceroses (*Ceratotherium simum*). *PLOS One*, *12*(12), e0187751.
- Waldram, M., Bond, W., & Stock, W. (2008). Ecological engineering by a mega-grazer: White rhino impacts on a South African savanna. *Ecosystems*, *11*(1), 101–112.
- Wolfensohn, S., Shotton, J., Bowley, H., Davies, S., Thompson, S., & Justice, W. (2018). Assessment of welfare in zoo animals: Towards optimum quality of life. *Animals*, *8*(7), 110–126.