

---

---

## **HYPERSENSITIVITY TO CAPTIVE DIETS AS A POSSIBLE UNDERLYING ETIOLOGY FOR CLINICAL SIGNS AND PATHOLOGY IN CAPTIVE BLACK RHINOCEROS (*Diceros bicornis*)**

*Ray L. Ball, DVM,\* Mike S. Burton, VMD, Genny Dumonceaux, DVM, and John H. Olsen, DVM*

*Busch Gardens Tampa Bay, 3605 Bougainvillea Drive, Tampa, FL 33612 USA*

Reprinted with permission of the Comparative Nutrition Society (CNS). 2006. Proc. Sixth Biennial Conference, Keystone, Colorado. Information in the CNS abstracts is not peer reviewed and cannot be considered endorsed by the society.

### **Abstract**

Black rhinoceros in captivity have been plagued by a host of clinical entities. These include superficial necrolytic dermatitis (SND), hemosiderosis, hemolytic and, non-hemolytic, anemia, and recently the idiopathic hemorrhagic vasculopathy syndrome (IHVS) has been described in a group of black rhinoceros.<sup>3,4</sup> Secondary infections are also noteworthy and include salmonellosis, leptospirosis, tuberculosis, and aspergillosis.<sup>4</sup> Information collected over the last several years have led to a theory that would potentially unite the pathologies and clinical conditions seen in captive black rhinoceros. A nutritional basis has long been suspected and the focus has tended to be on specific nutrients like iron or fatty acids.<sup>6,7</sup> Another possibility is that captive diets may be an antigenic source and initiate a cascade of events that may lead to the pathologies and clinical conditions encountered in captive black rhinoceros.

Figure 1 summarizes the proposed pathophysiology with dietary hypersensitivity as the inciting cause for health issues seen in captive black rhinoceros. Differences exist in the level of inflammation between captive and wild black rhinoceros as evidenced with ferritin levels. The differences between wild rhinoceros and captive rhinoceros in regards to ferritin are well documented.<sup>6</sup> While ferritin is used as a marker for tissue storage of iron, it is an acute phase inflammatory protein as well. Ferritin levels increase over time spent in captivity. This has been assumed to be to constant iron loading but a persistent inflammatory process could result in the same ferritin changes. A diet trial at Busch Gardens Tampa Bay (BGT) was undertaken in three adult male black rhinoceros (*Diceros bicornis michaeli*) (Studbook Nos. 518, 12-yr-old; 0786, 5-yr-old; 0864, 4-yr-old) in which a commercial browser pellet was substituted with a low starch, high physical effective fiber diet designed for giraffe. Basic hematology, serum chemistries, serum ferritin, and immune profiles were collected. Serum ferritin levels were checked (Kansas State Veterinary Diagnostic Laboratory, 1800 Dennison Avenue, D-117, Manhattan, Kansas 66506-5601 USA) at the beginning and ending of the trial and are listed in Table 2. The iron content of the new diet averages around 400 ppm but varies slightly between lots. The browser pellets contained 370 ppm iron. Ferritin does spike with any inflammatory process, including immobilizations. On one occasion, rhinoceros 518 was immobilized for electroejaculation. Serum ferritin on that procedure was 5466 ng/ml. One week later it returned to “baseline” of

---

---

2443ng/ml. Lymphocyte proliferation<sup>8</sup> was evaluated at Mote Marine Laboratory, (1600 Ken Thompson Parkway, Sarasota, Florida 34236 USA) at the beginning and end of the trial.<sup>8</sup> Concanavalin A (Con A) and phytohemagglutinin (PHA) were used as mitogens to stimulate lymphocyte proliferation. Immune response was slightly less at the end of the 5-mo time period. The difference may not be statistically significant but the clinical significance may be real given other inflammatory mediators had been reduced. Antiphospholipid antibodies (APhL) have not been evaluated as of this writing. A reduction in serum ferritin in spite of a higher iron diet suggests something other than iron intake is taking place here and a change in inflammation is suspected. Antiphospholipid antibodies (APhL) have been examined in captive and wild black rhinoceros, as well.<sup>1</sup> Wild black rhinoceros have lower levels of these antibodies compared to captive ones. A rising level can be seen when young captive rhinoceros are weaned onto solid foods.<sup>1</sup> This rise in APhL parallels that seen in ferritin. APhL are commonly seen in inflammatory processes in people. It is believed that in black rhinoceros they are reflective of an increased inflammatory stimulus in captivity. Given the various conditions wild rhinoceros are often in regards to parasites and wounds, a reasonable deduction would be the diet in captivity could be inciting the inflammation.

Support for a dietary source of gastrointestinal inflammation also comes from recent field work. Eleven black rhinoceros were recently translocated from Hluhluwe-Imfolozi Wildlife Park in South Africa. At capture all rhinoceros had fecals collected for various projects. Fecal hemacult were analyzed using a commercial kit (Hemacult®, Beckman Coulter, Inc., 4300 N. Harbor Blvd., Fullerton, California 92834-3100 USA) animal side for the presence of fecal occult blood. All eleven samples were negative. Fecal hemacult test are always positive in all species of rhinoceros at BGT. This test is not considered reliable in horses as the hindgut can readily degrade large amounts of hemoglobin, hence masking gastric bleeding.<sup>5</sup> While negative results can possibly be false negatives, a false positive seems very unlikely and suggest some bleeding in the gastrointestinal tract. Tannins or other porphyrin containing substances that could potentially interfere with this assay seem much more likely to occur in wild rhinoceros consuming natural browse material. A recent epidemiologic project looking at the health issues in captive black rhinoceros listed diarrhea as the most common problem seen.<sup>2</sup> Food allergen testing has also been conducted at a commercial veterinary food allergen testing facility (Bio-Medical Services, P.O. Box 26600, Austin, Texas 78755 USA, [www.bmslab.com](http://www.bmslab.com)) on captive black rhinoceros at BGT and five wild black rhinoceros from Zimbabwe. There is a fair amount of variability in the profiles between food items in the captive rhinoceros but corn and wheat are consistently reacting as antigens on the assay. A young captive born rhinoceros (61115) shows an increase in the level of reactivity to several items over time with a large increase occurring after weaning. Most interesting is the large differences between the wild rhinoceros (Zim 1-5) and the captive rhinoceros. Persistence of these two inflammatory proteins, ferritin and APhL, may lead to problems directly. Antiphospholipid can cause microthrombi and mimic problems seen in captive black rhinoceros.<sup>1</sup> The problems with hemosiderosis are well documented in black rhinoceros.<sup>6</sup>

---



---

## ACKNOWLEDGMENTS

We would like to thank the Cathy Walsh at Mote Marine, Drs. Dave Cooper, Markus Hofmeyer, and Peter Buss in South Africa, Mary Port and Heather hHenry at the Veterinary Hospital at BGT, and Jason Green , Kristin Forker, and Derek Weatherford and the entire rhinoceros crew at BGT.

## LITERATURE CITED

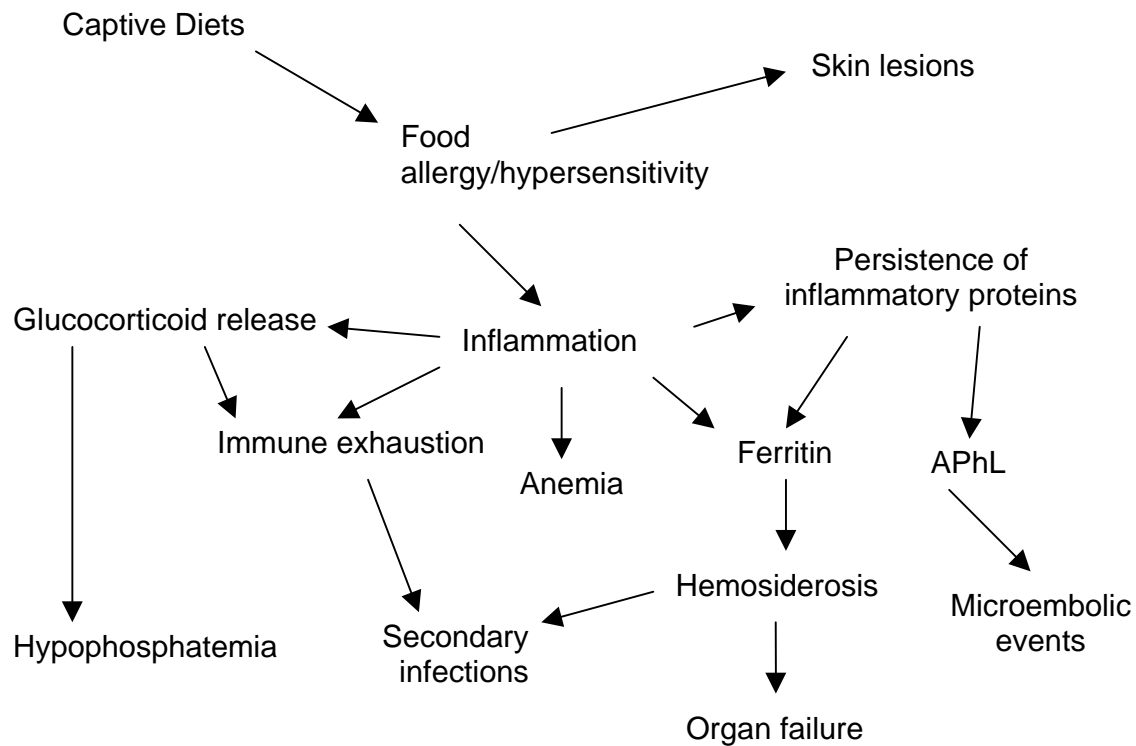
1. Ball, R.L., M. Morrow, P.M. Hofmeyer, and P. Buss. 2005. Comparison of anti-phospholipid antibodies between wild and captive black rhinoceros (*Bicornis bicornis*): Implications for health and repatriation. Proc. Inst. Zoo Wildl. Res. Pp.72-74.
2. Dennis, P., W.L. Saville, E.S. Blumer, et al. 2003. Preliminary review of an epidemiological survey of black rhinoceros in captivity in the United States. Proc. Am. Assoc. Zoo Vet. Annu. Meet. Pp. 247.
3. Kock, N., C. Foggin, M. Kock, and R. Kock. 1992. Hemosiderosis in the black rhinoceros (*Diceros bicornis*): comparison of free-ranging and recently captured with translocated and captive animals. J. Zoo Wildl. Med. 23:230-234.
4. Miller, R.E., 2003. Rhinocerotidae. In: M. Fowler and R.E.Miller (eds.): Zoo and Wildlife Medicine, 5th Ed. Pp. 563-66.
5. Murray, M. 1997. The Gastrointestinal System. In: Robinson, N.E. (ed.). Current Therapy in Equine Medicine. W.B. Saunders Co., Philadelphia, PA. Pp. 169.
6. Paglia, D. and P. Dennis. 1999. Role of chronic iron overload in multiple disorders of captive black rhinoceros (*Diceros bicornis*). Proc. Am. Assoc. Zoo Vet. Annu. Meet. Pp. 163-171
7. Suedmeyer, W.K. and E.S. Dierenfeld. 1998. Clinical experience with fatty acid supplementation in a group of black rhinoceros (*Diceros bicornis*) Proc. Am. Assoc. Zoo Vet. Annu. Meet. Pp. 113-115.
8. Vance C.K., S. Kennedy-Stoskopf, A.R. Obringer, and T.L. Roth. 2004. Comparative studies of mitogen- and antigen-induced lymphocyte proliferation in four captive rhinoceros species. J. Zoo Wildl. Med. 34(4):435-446.

**Table 1.** Food item hypersensitivity profiles on wild and captive black rhinoceros. (N=Negative ≤150, BL=Borderline 151-174, BL-P=Borderline-Positive 175-199, P=Positive 200-400, HP=Highly Positive >400). Dates from wild rhinoceros are approximates.

	518	0786	0786	0786	0786	Zim1	Zim2	Zim3	Zim4	Zim5
	07/12/	12/21/	10/22/	02/28/	12/14/	04/01/	04/01/	04/01/	04/01/	04/01/
Sample Date	01	00	04	05	05	01	01	01	01	01
Barley	129	100	237	256	261	100	134	100	100	111
Soy Beans	140	164	149	176	196	129	100	138	100	135
Corn	159	138	237	214	239	149	243	165	165	177
Wheat	163	100	253	276	357	127	165	100	127	105
Brown Rice	146	101	124	165	219	100	100	118	100	100
Potato	102	132	102	100	100	100	100	100	100	125
Beets	112	105	228	157	155	100	108	100	194	134
Carrot	117	100	149	142	166	100	100	100	100	100
Sweet										
Potato	121	100	100	105	191	100	148	100	102	100
Yeast	100	100	100	100	100	100	100	101	100	100
Milk/Cow's	100	100	100	100	100	100	126	100	100	100

**Table 2.** Beginning and ending serum ferritin (ng/ml) from diet trial on three adult male captive black rhinoceros.

	Jan-05	Aug-05
518	4405	2490
786	10733	2860
864	12671	6052



**Figure 1.** Proposed pathophysiology involving dietary hypersensitivity and gastrointestinal inflammation for the clinical and pathologic conditions in captive black rhinoceros.