

M. D. Kock and M. W. Atkinson. Intensive management of an endangered species: Long-term effects of chemical immobilisation and dehorning on health and reproduction in a discrete population of black rhinoceroses in Zimbabwe. *Proceedings, Wildlife Disease Association Conference, Fairbanks, Alaska, July 1996*, p. 38.

The paper discusses a population of black rhinoceroses that were immobilised for horn removal or attachment of radio collars in the Sinamatella Intensive Protection Zone (IPZ) in 1992. At that time the population was estimated at >60 animals, and despite deaths in 1993 and 1994, the 1996 population estimate is >65 animals. The paper discusses the long-term effects of intervention strategies in the survival of this species, and concludes that intervention had minimal impact on the survival of this population.

N. D. Kock, C. Foggin and M. D. Kock. Pathological findings in Zimbabwean black rhinoceroses (*Diceros bicornis*): A summary. *Proceedings, Wildlife Disease Association Conference, Fairbanks, Alaska, July, 1996*, p. 39.

The pathological findings from over eight years of investigation of natural deaths among free-ranging and translocated black rhinoceroses are presented and discussed. Findings include haemosiderosis, as an indication of haemolysis, coronary artery aneurysm of unknown aetiology, septicaemia due to *Streptococcus equisimilis* secondary to infected wounds, salmonellosis, degenerative arthritis, ulcerative skin lesions of parasitic aetiology (*Stefanofilaria* spp.), and a report of endoparasites recovered from faecal examination.

N. D. Kock, F. Jongejan, A. H. M. van Vliet and K. Charlton. Evidence of *Cowdria ruminantium* infection in wildlife species in Zimbabwe. *Proceedings, Wildlife Disease Association Conference, Fairbanks, Alaska, July, 1996*, p. 71.

Sera from free ranging Zimbabwean black (*Diceros bicornis*) and white (*Ceratotherium simum*) rhinoceroses tested positive for antibodies to *Cowdria ruminantium* by monoclonal antibody-mediated competitive enzyme-linked immunosorbent assay. However, as with other serological tests for *C. ruminantium*, the possibility of cross reactions with *Ehrlichia* spp. had to be considered. A more specific polymerase chain reaction assay was developed for the detection of *C. ruminantium* DNA in blood and bone marrow, and tsessebe (*Damaliscus lunatus*), waterbuck (*Kobus ellipsiprymnus*), impala (*Aepyceros melampus*) and reedbuck (*Redunca arundinum*) were tested. A 297 base pair segment of DNA derived from the *C. ruminantium* MAP1 gene was amplified from blood and bone marrow of tsessebe, from blood of waterbuck and from bone marrow of impala. Results were confirmed by southern blot hybridisation.

INSTRUCTIONS TO AUTHORS

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Numbers up to nine should be written in full unless followed by an abbreviated unit of measurement. Numbers above nine should be given in figures unless used to begin a sentence. All measurements should be in the SI metric system and abbreviations should conform to common usage. Abbreviated names may be used in the text when the original name followed by the abbreviation in parentheses has been used once. Drugs should be referred to by their pharmacological names, as found in the British Veterinary Codex or other pharmacopoeias. Where the trade name is more commonly known this should be given in parentheses with the name of the manufacturer.

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The paper should be headed with a short informative title followed by the name(s) of the author(s). If any author is female one given name should be stated in full. This should be followed by the address of the authors at which the work was performed. If the present address of any author has changed it may appear as a footnote to the first page of the article.

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6. Acknowledgements of assistance.